

The diversity of annelids in subterranean waters: a case study from Poland

Elzbieta Dumnicka^{1,*}, Joanna Galas¹, Mariola Krodkiewska² and Agnieszka Pociecha¹

¹ Institute of Nature Conservation, Polish Academy of Sciences, A. Mickiewicza 33, 31-120 Kraków, Poland

² Institute of Biology, Biotechnology and Environmental Protection, University of Silesia, Bankowa 9, 40-007 Katowice, Poland

Received: 9 December 2019 / Accepted: 21 February 2020

Abstract – Not all invertebrate groups commonly occur in subterranean waters but annelids live in surface and underground habitats. The annelid species' richness in various underground waters (wells and interstitial and cave waters) and surface streams of Poland was compared, and the habitat preferences for the most frequent species were determined. Until now, 111 annelid taxa (mainly oligochaetes) had been identified in underground waters in Poland, with higher numbers (71) in the interstitial habitat than in stream bottoms (62). The number of species identified in the caves and wells was distinctly lower (54 and 29, respectively). The Correspondence Analysis did not separate the samples from various underground water types into distinct groups, and the distribution of well fauna was especially scattered (in the ordination diagram) because abiotic parameters differ strongly in studied wells. Only three stygobiontic species (*Cernosvitoviella parviseta*, *Enchytraeus dominicae* and *Trichodrilus moravicus*) were related to some caves. The analysis of the available data indicate that to obtain a comprehensive picture of the aquatic fauna in a given country all types of subterranean aquatic habitats should be sampled and taken into account. Moreover, to ascertain the composition of benthic invertebrates in running waters, investigation of the interstitial habitat should also be performed.

Keywords: Oligochaetes / interstitial waters / wells / cave waters / stream benthos

Résumé – La diversité des annélides dans les eaux souterraines : une étude de cas en Pologne.

Tous les groupes d'invertébrés ne sont pas présents dans les eaux souterraines, mais les annélides vivent dans des habitats de surface et souterrains. La richesse des espèces d'annélides dans les différentes eaux souterraines (puits, eaux interstitielles et grottes) et les cours d'eau de surface de Pologne a été comparée, et les préférences d'habitat des espèces les plus fréquentes ont été déterminées. Jusqu'à présent, 111 taxons d'annélides (principalement des oligochètes) ont été identifiés dans les eaux souterraines en Pologne, avec un nombre plus élevé (71) dans l'habitat interstitiel que dans le fond des cours d'eau (62). Le nombre d'espèces identifiées dans les grottes et les puits était nettement plus faible (54 et 29, respectivement). L'analyse des correspondances n'a pas séparé les échantillons des différents types d'eaux souterraines en groupes distincts, et la distribution de la faune des puits était particulièrement dispersée (dans le diagramme d'ordination) car les paramètres abiotiques diffèrent fortement dans les puits étudiés. Seules trois espèces stygobiontiques (*Cernosvitoviella parviseta*, *Enchytraeus dominicae* et *Trichodrilus moravicus*) étaient apparentées à certaines grottes. L'analyse des données disponibles indique que pour obtenir une image complète de la faune aquatique dans un pays donné, tous les types d'habitats aquatiques souterrains doivent être échantillonnés et pris en compte. En outre, pour déterminer la composition des invertébrés benthiques dans les eaux courantes, il convient également de procéder à une étude de l'habitat interstitiel.

Mots clés : oligochètes / eaux interstitielles / puits / eaux souterraines / benthos de rivière

*Corresponding author: dumnicka@iop.krakow.pl

1 Introduction

Subterranean waters constitute the majority of freshwater resources, but invertebrates inhabit only its relatively shallow level, primarily in unconfined aquifers (Gibert *et al.*, 1994a). These invertebrates can be accessible in caves, springs, and dug and drilled wells. Springs are outflows of underground waters, and typical subterranean species such as stygobionts (obligate subterranean aquatic organisms) can sometimes be found in them (Dumnicka and Galas, 2017). However, due to the character of this biotope, which is similar to other surface water bodies, they are inhabited by the benthic community (along with some species specific for springs). For this reason, springs were not taken into consideration in this paper. Shallow subterranean habitats such as interstitial waters (Juberthie and Decu, 1994; Culver and Papan, 2009) as well as hypotelminorheic and epikarst waters (Culver and Papan, 2011) and alluvial mesovoid shallow substratum (Ortuño *et al.*, 2013) also belong to the category of subterranean waters, but the fauna of both these aforementioned habitats was not studied in Poland.

Groundwater fauna consists of: stygobionts – organisms reproducing exclusively in this habitat, stygophiles – species completing their life cycle in both subterranean and surface waters, and stygoxens – species that can accidentally be found in underground water (Pacioglu 2010). These groups have different degrees of adaptation to groundwaters (Gibert *et al.*, 1994b), but even stygobionts can be found occasionally in surface waters (Krzanowski *et al.*, 1965; Dumnicka *et al.*, 2018). Composition of invertebrate benthic fauna in surface and subterranean waters differs significantly: non-insect species (mainly crustaceans, water mites and annelids) are the most abundant and diverse in underground waters (*e.g.*, Sket, 1999; Košel, 2009; Martin *et al.*, 2009; Chertoprud *et al.*, 2016), while insect fauna is usually rich in the benthos of non-polluted stagnant and running surface waters of Europe (Starmach *et al.*, 1976; Dijkstra *et al.*, 2014). The differences between the composition of bottom and interstitial macro-invertebrate fauna are often small (Mary and Marmonier, 2000; Meleg *et al.*, 2009), but the larvae of insects are sometimes scarce in the interstitial habitat (Danielopol, 1984; Creuzé des Châtelliers *et al.*, 1992).

In Poland, one of the most studied groups living in subterranean waters (including small, temporary pools occurring in caves, principally in the Kraków-Częstochowa Upland) are annelids. Studies on this taxonomic group started at the end of the 19th century when Jaworowski (1893) found a few species in municipal wells in Kraków. Only eighty years later, Kasprzak (1973a,b) published new data from a number of wells situated in various regions of Poland. More extensive studies were conducted by Dumnicka (2000) and Dumnicka *et al.* (2017) on oligochaetes in this habitat. The first information about the presence of oligochaetes in caves was published by Demel (1918), who found only a singular lumbricid taxon. From 30 yr in the 20th century, cave annelids were studied by Pax & Maschke (1935), Moszyński (1936) and Stammer (1936) in the Sudetes, whereas Skalski (1967), Kasprzak (1973b) and Kasprzak & Zajonc (1980) published information about this group from singular caves located in the Tatra Mountains. The majority of data concerning oligochaetes from many Polish caves were elaborated on by Dumnicka and

Table 1. Number of analysed objects and number of publications concerning particular subterranean habitats. In some papers, results obtained from two or three different habitats were included.

Habitat	Number of studied objects	Number of publications
Wells	circa 70	7
Caves	31	24
Interstitial waters	46	8
Water works	2	2

co-workers (Dumnicka, 2000 (in this paper 11 previous publications of this author were cited) Dumnicka, 2003, 2009; Dumnicka *et al.*, 2015, 2016). A few data sets available from artificial subterranean spaces such as adits or quarries (Moszyński 1936; Dumnicka 1996) are included with data from caves in this paper. The studies on annelid fauna of interstitial waters began in the second half of the 20th century, but this habitat was intensively investigated in following years (Kasprzak, 1973a,b,c; 1975, 1976, 1979a,b; Dumnicka, 2000; Dumnicka *et al.*, 2018). In Poland, very few studies dealing with annelids were conducted in the waterworks systems (Moszyński, 1934; Pawłowski, 1951) directly connected with underground waters, which are sometimes inhabited by numerous and diverse invertebrates (Łuczak *et al.*, 1980).

Aims:

The present study summarizes all available published information on annelids inhabiting subterranean aquatic habitats in Poland in order to determine (1) whether there is a difference in annelid composition among different habitats; and (2) what factors influence annelid species richness and their habitat references

2 Material and methods

In this study data from 38 papers concerning oligochaetes studied in four subterranean habitats have been used (Tab. 1, Fig. 1). Sometimes a few sampling stations were studied in the same cave, this also applies to particular running waters. The majority of papers dealing with oligochaeta composition from subterranean waters resulted from qualitative samples collected using a bottom scraper. Much less frequently quantitative samples have been collected using the Ekman sampler or a bottom scraper. In both the methods, the collected sediments were washed using nets with various mesh size. Fauna samples from interstitial waters were collected from holes (40–50 cm deep), which were dug in sediments on the river border (Karaman-Chappuis method). Specimens from samples of all types were sorted using a stereoscopic microscope or with naked eyes. Especially in previous papers, methods are not strictly described. From various localities samples were collected once or several times.

In analysed papers usually the total number of collected specimens or their relative abundance *e.g.* “high number”, “numerous”, “fairly numerous” *etc.* or the relative scale was used (Tab. 2). In Table 3, the number of records represents the number of species findings in particular subterranean aquatic

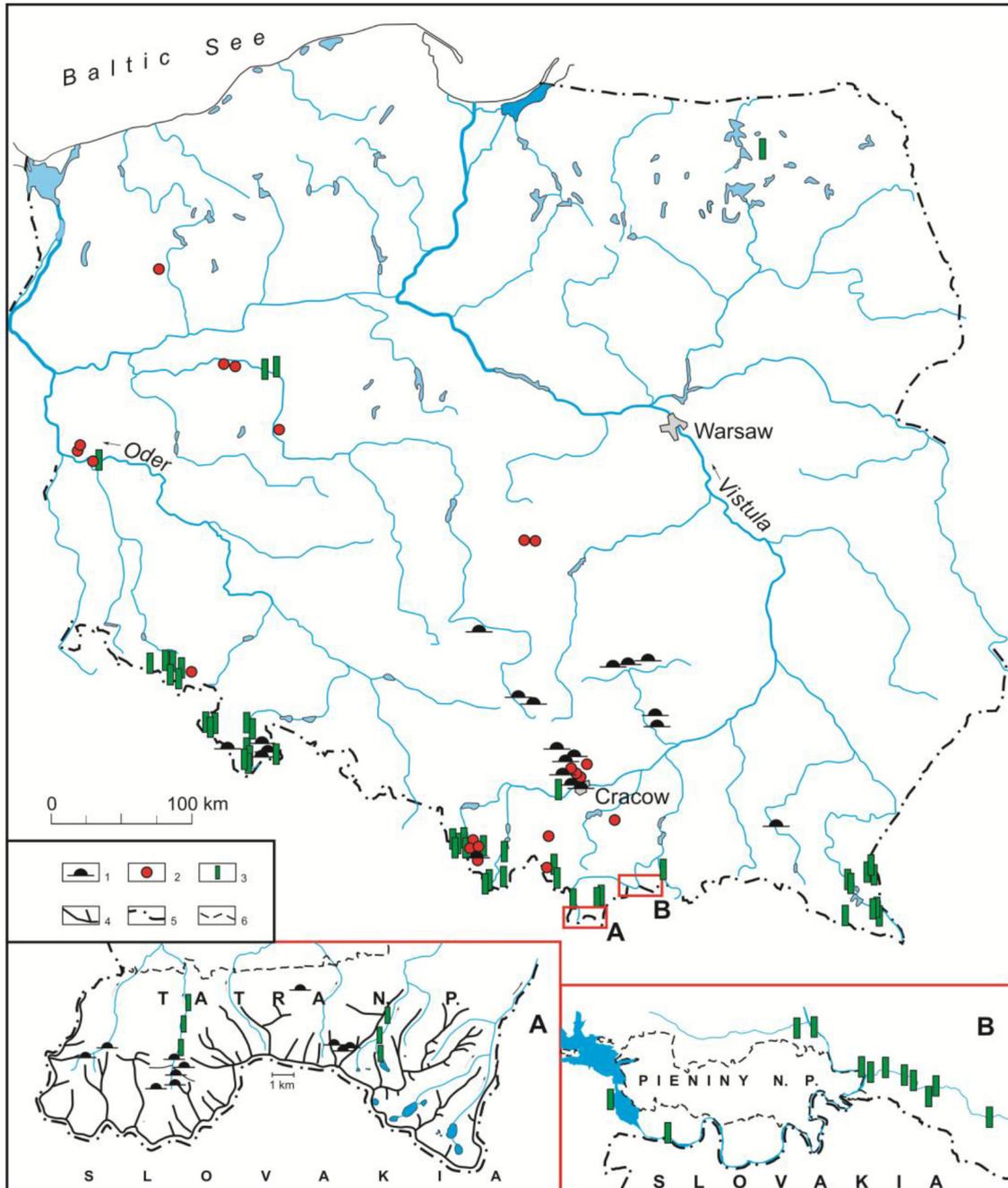


Fig. 1. The map of Poland showing the distribution of studied habitats; the areas of the Tatra Mts (A) and Pieniny Mts (B) are enlarged. 1–caves, 2–wells, 3–interstitial waters, 4–mountain ranges, 5–country border, 6–national park border.

Table 2. Relative abundance description sheet for Annelida taxa.

Description	Relative abundance scale
Single individuals	1
Low abundance	2
Medium abundance	3
Abundant	4
Very abundant	5

habitats. The samples without annelids were not taken into account.

Systematics of Annelida according to Fauna Europaea (www.fauna-eu.org) as well as Kahl and Pilipiuk (2004) was used.

2.1 Data analysis

In statistical analyses, the waterworks were not included because the data came only from two localities (the municipal

Table 3. Number of annelid species in various subterranean habitats. In streams benthic and interstitial annelids fauna was studied.

Taxa	Bottom sedim.	Subterranean habitats				
		Σ records	Interst. waters	Cave waters	Wells	Waterworks device
Polychaeta						
<i>Hrabeiella periglandulata</i> Pižl & Chalupsky, 1984		1		1		
<i>Troglochaetus beranecki</i> Delachaux, 1921		2		2		
Aphanoneura						
<i>Potamodrilus fluviatilis</i> Lastočkin, 1935	1	1	1			
<i>Aeolosoma</i> sp. Ehrenberg, 1831		5		4	1	
<i>Aeolosoma niveum</i> Leydig, 1865		1				1
<i>Aeolosoma tenebrarum</i> Vejdovsky, 1880		1	1			
Oligochaeta						
<i>Propappus volki</i> Michaelsen, 1916	22	26	21	5		
<i>Achaeta eiseni</i> Vejdovsky, 1877		2		2		
<i>Achaeta seminalis</i> Kasprzak, 1972		1	1			
<i>Bryodrilus ehlersi</i> Ude, 1892		2		2		
<i>Buchholzia appendiculata</i> (Buchholz, 1862)	2	8	3	5		
<i>Cernosvitoviella atrata</i> (Bretscher, 1903)	3	13	5	7	1	
<i>Cernosvitoviella carpatica</i> Niel. & Christ., 1959	1	1		1		
<i>Cernosvitoviella minor</i> Dózsa-Farkas, 1990		1		1		
<i>Cernosvitoviella parviseta</i> Gadzinska, 1974		6		4	2	
<i>Cernosvitoviella tatrensis</i> (Kowalewski, 1916)	3	9	3	6		
<i>Cognettia anomala</i> (Cernosvitov, 1928)	1					
<i>Cognettia glandulosa</i> (Michaelsen, 1888)	2	3	2	1		
<i>Cognettia sphagnetorum</i> (Vejdovsky, 1877)	3	10	5	4	1	
<i>Enchytraeus albidus</i> Henle, 1837		1			1	
<i>Enchytraeus buchholzi</i> Vejdovsky, 1879	2	24	5	13	6	
<i>Enchytraeus christenseni</i> Dózsa-Farkas, 1992		1	1			
<i>Enchytraeus dominicae</i> Dumnicka, 1976		14	1	11	2	
<i>Enchytraeus lacteus</i> Niel. & Christ., 1961		5		2	3	
<i>Enchytraeus mariae</i> Kasprzak, 1973		1		1		
<i>Enchytraeus norvegicus</i> Abrahamsen, 1969	1	1	1			
<i>Enchytraeus polonicus</i> Dumnicka, 1977		2		2		
<i>Fridericia bisetosa</i> (Levinsen, 1884)	1					
<i>Fridericia bulbosa</i> (Rosa, 1887)	1	5	2	3		
<i>Fridericia galba</i> (Hoffmeister, 1843)	2	3	2	1		
<i>Fridericia leydigi</i> Vejdovsky, 1877	1	1	1			
<i>Fridericia maculata</i> Issel, 1904		1		1		
<i>Fridericia paroniana</i> Issel, 1904	1	1	1			
<i>Fridericia perrieri</i> (Vejdovsky, 1877)		1	1			
<i>Fridericia ratzeli</i> (Eisen, 1872)	2	3	2	1		
<i>Fridericia semisetosa</i> Dózsa-Farkas, 1970		1		1		
<i>Fridericia tubulosa</i> Dózsa-Farkas, 1972	1	1	1			
<i>Henlea gubleri</i> Bretscher, 1903		1		1		
<i>Henlea nasuta</i> (Eisen, 1878)	1	2		2		
<i>Henlea perpusilla</i> Friend, 1911	1	7	1	6		
<i>Henlea similis</i> Niel. & Christ., 1959	2	2	2			
<i>Henlea ventriculosa</i> (d'Udekem, 1854)	3	5	3	1	1	
<i>Lumbricillus rivalis</i> (Levinsen, 1884)	2	2	2			
<i>Marionina argentea</i> (Michaelsen, 1889)	2	16	2	12	2	
<i>Marionina libra</i> Niel. & Christ., 1959		1	1			
<i>Marionina riparia</i> Bretscher, 1899		8	1	7		
<i>Marioniana spicula</i> (Leuckart, 1847)		1	1			
<i>Mesenchytraeus armatus</i> (Levinsen, 1884)	3	4	3	1		
<i>Mesenchytraeus sanguineus</i> Niel. & Christ., 1959		1	1			
<i>Amphichaeta leydigi</i> Tauber, 1879		1		1		
<i>Aulophorus furcatus</i> (Oken, 1815)		1	1			
<i>Chaetogaster diaphanus</i> (Gruith., 1828)	1	2	1	1		

Table 3. (continued).

Taxa	Bottom sedim.	Subterranean habitats				
		Σ records	Interst. waters	Cave waters	Wells	Waterworks device
<i>Chaetogaster diastrophus</i> (Gruith., 1828)	1	2	1	1		
<i>Chaetogaster langi</i> Bretscher, 1896		1				1
<i>Nais alpina</i> Sperber, 1948	2	3	2		1	
<i>Nais barbata</i> O.F. Muller, 1774	1	1	1			
<i>Nais bretscheri</i> Michaelsen, 1899	1	3	3			
<i>Nais communis</i> Piguet, 1906	6	12	6	3	3	
<i>Nais elinguis</i> O.F. Muller, 1774	14	17	14	2		1
<i>Nais pardalis</i> Piguet, 1906	2	2	2			
<i>Nais pseudobtusa</i> Piguet, 1906	2	2	2			
<i>Nais simplex</i> Piguet, 1906	2	3	2		1	
<i>Nais variabilis</i> Piguet, 1906	1	3	1	1		1
<i>Ophidonais serpentina</i> (O.F. Muller, 1774)		1	1			
<i>Pristina bilobata</i> (Bretscher, 1903)	5	5	5			
<i>Pristina aequiseta</i> Bourne, 1891	29	33	29		3	1
<i>Pristina jenkiniae</i> Stephenson, 1932		1		1		
<i>Pristina longiseta</i> Ehrenberg, 1828	1	4	4			
<i>Pristina menoni</i> Aiyer, 1929	29	32	29	1	2	
<i>Slavina appendiculata</i> (d'Udekem, 1855)		2	1		1	
<i>Vejdovskyella intermedia</i> (Bretscher, 1896)		1		1		
<i>Aulodrilus pluriseta</i> (Piguet, 1906)	1	1	1			
<i>Edukemius benedi</i> (d'Udekem, 1855)	3	4	3		1	
<i>Gianius aquaedulcis</i> (Hrabě, 1960)	1					
<i>Haber zavreli</i> (Hrabě, 1942)		3			3	
<i>Ilyodrilus templetoni</i> (Southern, 1909)	1	1	1			
<i>Limnodrilus</i> sp. juv. Claparède, 1862		1		1		
<i>Limnodrilus claparedeanus</i> Ratzel, 1869	2	2	2			
<i>Limnodrilus hoffmeisteri</i> Claparède, 1862	8	8	8			
<i>Limnodrilus udekemianus</i> Claparède, 1862	1	2	2			
<i>Epirodilus pygmaeus</i> (Hrabě, 1935)	2	2	2			
<i>Potamothrix hammoniensis</i> (Michaelsen, 1901)	1	2	2			
<i>Potamothrix moldaviensis</i> Vejd. & Mrazek, 1903	1	2	1		1	
<i>Psammoryctides albicola</i> (Michael., 1901)	3	3	3			
<i>Psammoryctides barbatus</i> (Grübe, 1861)	1	1	1			
<i>Rhyacodrilus coccineus</i> (Vejdovsky, 1876)	4	7	6		1	
<i>Rhyacodrilus falciformis</i> Bretscher, 1901		3		3		
<i>Rhyacodrilus subterraneus</i> Hrabě, 1963		2	1		1	
<i>Spirosperma ferox</i> (Eisen, 1879)	3	4	4			
<i>Tubifex ignotus</i> (Štolc, 1886)		1		1		
<i>Tubifex tubifex</i> (O.F. Muller, 1774)	6	10	6	2	2	
<i>Lumbriculus variegatus</i> (O.F. Muller, 1774)	12	14	12	2		
<i>Rhynchelmis</i> sp. juv. Hoffmeister, 1843		1	1			
<i>Stylodrilus brachystylus</i> Hrabě, 1929	1	4	3		1	
<i>Stylodrilus heringianus</i> Claparède, 1862	16	19	16	3		
<i>Stylodrilus parvus</i> (Hrabě & Cernosvitov, 1927)		1		1		
<i>Trichodrilus</i> sp. juv. Claparède, 1862		1			1	
<i>Trichodrilus cernosvitovi</i> Hrabě, 1937	1	2	2			
<i>Trichodrilus moravicus</i> Hrabě, 1937	2	7	2	4	1	
<i>Trichodrilus pragensis</i> Vejdovsky, 1876		2		2		
<i>Trichodrilus spelaeus</i> Moszyński, 1936		1		1		
<i>Haplotaxis gordioides</i> (Hartmann, 1821)	18	25	18	4	3	
<i>Dendrobaena</i> sp. Eisen, 1874		1		1		
<i>Dendrodrilus rubidus</i> (Savigny, 1826)		2		2		
<i>Eisenia fetida</i> (Savigny, 1826)		1			1	
<i>Eiseniella tetraedra</i> (Savigny, 1826)	1	3	1	1	1	
<i>Lumbricus terrestris</i> L., 1758		1			1	

Table 3. (continued).

Taxa	Bottom sedim.	Subterranean habitats				
		Σ records	Interst. waters	Cave waters	Wells	Waterworks device
Hirudinea						
<i>Erpobdella octoculata</i> (L., 1758)		1				1
<i>Erpobdella nigricollis</i> (Brandes, 1900)		1				1
<i>Glossiphonia complanata</i> (L., 1758)		1				1
<i>Helobdella stagnalis</i> (L., 1758)		1				1
Σ	250	489	280	151	49	10

waterworks in Warsaw and Poznań cities). An indirect analysis (a correspondence analysis), based on the relative abundance of annelid species, was carried out using CANOCO for Windows version 4.5 (Ter Braak and Šmilauer, 2002) in order to inspect the distribution of species in the different underground water types. An indirect analysis (a correspondence analysis) was also calculated to estimate the distribution of oligochaetes in interstitial water and bottom sediments in the studied rivers. Only the individuals identified to a species level were included in the data matrix. Species in which the frequency was less than 5% were removed from the analysis to reduce noise in the data set. The results from the analysis were displayed graphically in an ordination diagram using the software program CanoDraw ver. 4.12.

Differences in the number of oligochaete species between the studied types of underground water were tested using the Kruskal–Wallis ANOVA and a multiple comparison post hoc test because the biological data did not reveal a normal distribution. The significance of the difference in the number of oligochaete species between interstitial water and bottom sediments in rivers was evaluated using the Student's *t*-test. The Pearson chi-square test was used to assess whether the frequency of the stygobiontic species occurrence in the compared types of underground waters deviated from a random pattern. The analysis could only be carried out for *Enchytraeus dominicae*, as the remaining stygobiontic species were present in too few samples. All calculations were performed using the software program Statistica for Windows ver. 13.1.

Species accumulation curves were calculated and constructed using the incidence-based coverage estimator (ICE) and S_{obs} (calculated according to the Mao Tao function procedures (Colwell, 2013)). The calculation was computed using Estimate S version 9.1. 0 for Windows with randomization 100 times without replacement.

3 Results

Investigations of annelid fauna in the subterranean habitats of Poland were performed mainly in the southern part of country and concentrated in the Tatra and Pieniny Mts. (Fig. 1). The number of studies conducted in central and northern part of Poland (where the caves are generally absent) was distinctly lower. Only about ten papers (from 38) deal with subterranean waters of Central and Northern parts of Poland where the caves are generally absent.

Until now, 111 annelid taxa have been found in subterranean waters of Poland, with many species found exclusively in one or two localities (Tab. 3). The species accumulation curves for the studied underground localities did not reach a plateau but continued to increase (as evidenced by the curve for the S_{obs}), indicating that not all annelid species were collected during sampling (Fig. 2).

Freshwater- and soil-dwelling Polychaeta are represented in Poland by only two species, both known from the underground waters (Fig. 3). From 13 species of Aphanoneura, only four taxa have been found in the subterranean environment, while Oligochaeta were represented by 100 taxa among 216 aquatic and terrestrial species known from Poland. Only in the Lumbriculidae family all species known from Poland were found in subterranean waters while in surface habitats (Fig. 3) stygobionts (four species) are missing. In other families, various numbers of species were identified in underground habitats: Tubificidae occurred at almost 60%, Naididae and Enchytraeidae at approximately 48% and 44% respectively, along with the smallest number from the family Lumbricidae, which accounted for approximately 16% of species. Leeches were absent in the wells, interstitial and cave waters; only four species belonging to two families from this group (Fig. 3) have been found in municipal waterworks. Among five classes of Annelida, only Branchiobdellea were not found in the subterranean environment in Poland.

The most numerous records (280) were collected in interstitial waters (Tab. 3), and the highest number of species (71) was found in this habitat as well. Benthic annelids, which were studied parallel to interstitial fauna, were found to have only 62 species present; however, the differences related to the mean species number found in these habitats were statistically insignificant ($t=4.678571$; $df=110$; $p > 0.05$). The correspondence analysis (CA) did not indicate a division of the samples from interstitial waters and bottom sediments in the studied rivers – these samples overlap in the graph in most cases (Fig. 4). The number of species found in the caves and wells is distinctly lower (54 and 29 species, respectively) than that from the interstitial habitat (Tab. 3). Similarly, the number of records from these habitats is lower than that from interstitial waters. The Kruskal–Wallis ANOVA test revealed statistically significant differences in the median number of species between the types of underground waters ($H=13.49893$; $p=0.0012$). However, multiple comparison post hoc tests showed statistically significant differences in the median number of species only between interstitial waters and

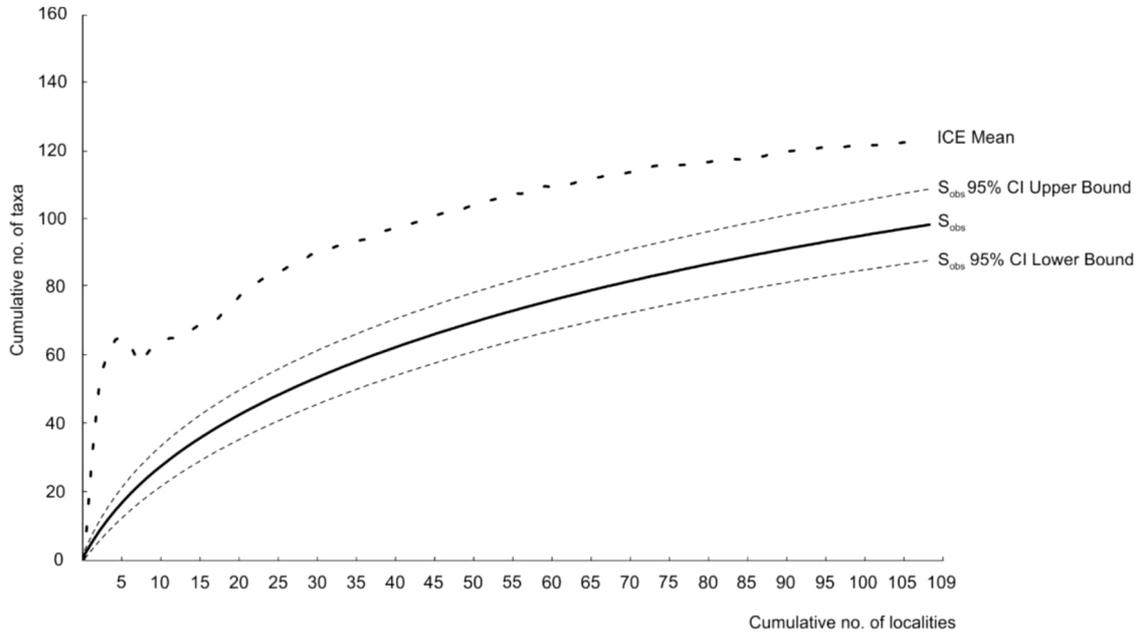


Fig. 2. Species accumulation curves and sample-based species richness estimator for study underground localities (S_{obs} – the total number of taxa in the polled samples calculated according to the Mao-Tau function, 95% CI – 95% confidence limit for S_{obs} , ICE mean – incidence-based coverage estimator).

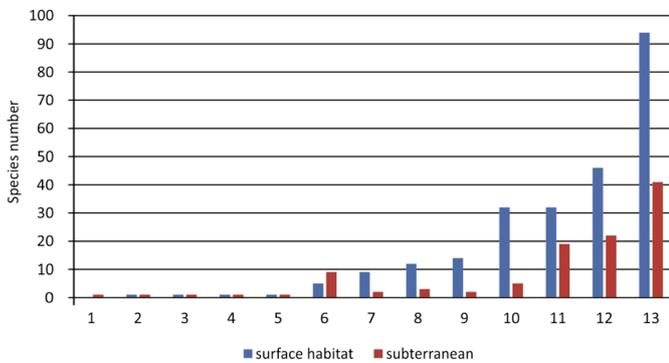


Fig. 3. Number of species (from various families) found in surface (blue bars) vs. underground (red bars) habitats: 1–Nerillidae, 2–Parergodrilidae, 3–Potamodrilidae, 4–Propappidae, 5–Haplotaxidae, 6–Lumbriculidae, 7–Erpbodellidae, 8–Aeolosomatidae, 9–Glossiphoniidae, 10–Lumbricidae, 11–Tubificidae, 12–Naididae, 13–Enchytraeidae.

two other underground water types (caves and wells). Only nine annelid species have been found in waterworks systems, but this habitat was studied sporadically (with 10 records).

The correspondence analysis (CA), conducted on the grounds of the relative abundance of oligochaete species, did not separate the samples of the studied underground water types into distinct groups (Fig. 5). In particular, the samples from wells are not clustered and are scattered on the diagram. The portion of samples from interstitial waters, located on the left side of the first axis in the graph, forms a separate group. It consists of species such as *Limnodrilus hoffmeisteri*, *Propappus volki*, *Stylodrilus heringianus*, *Nais elinguis*, *Haplotaxis*

gordioides, *Lumbriculus variegatus*, *Pristina menoni*, *P. aequiseta*, *Rhyacodrilus coccineus* and *Tubifex tubifex*. However, three stygobiontic species (*Cernosvitoviella parviseta*, *Enchytraeus dominicae* and *Trichodrilus moravicus*) (situated in the lower right side of the diagram) are related to some caves (Fig. 5), but only the occurrence of *E. dominicae* was more frequent in the caves (Pearson $\chi^2=18.98027$; $df=3$; $p < 0.001$). The results of the CA showed that the first two ordination axes explained 24.8% of the variability in the species data from the studied underground water types.

4 Discussion

According to Kahl and Pilipiuk (2004), 280 annelid species are known in Poland, and almost 40% of them have been found in subterranean waters. The species accumulation curves for the studied underground localities did not reach a plateau (as evidenced by the curve for the S_{obs}) indicating that not all annelid species were collected during sampling. Future studies should be primarily conducted in wells, since these structures reach subterranean waters in various geological layers, what is especially important in the regions without caves. In Poland, these areas are the least studied, particularly in the northern part of the country (Dumnicka and Galas, 2017).

A substantial number of species were found in a single or only a few localities, but the group of such species is heterogeneous. First, the majority of stygobionts are known from one or only a few localities in Poland (Dumnicka and Galas, 2017) besides numerous studies conducted in interstitial and cave waters. A good example is *Troglochaetus beranecki*, which was only found by Stammer (1936) in the Kłodzko Basin and was never found again despite intensive studies

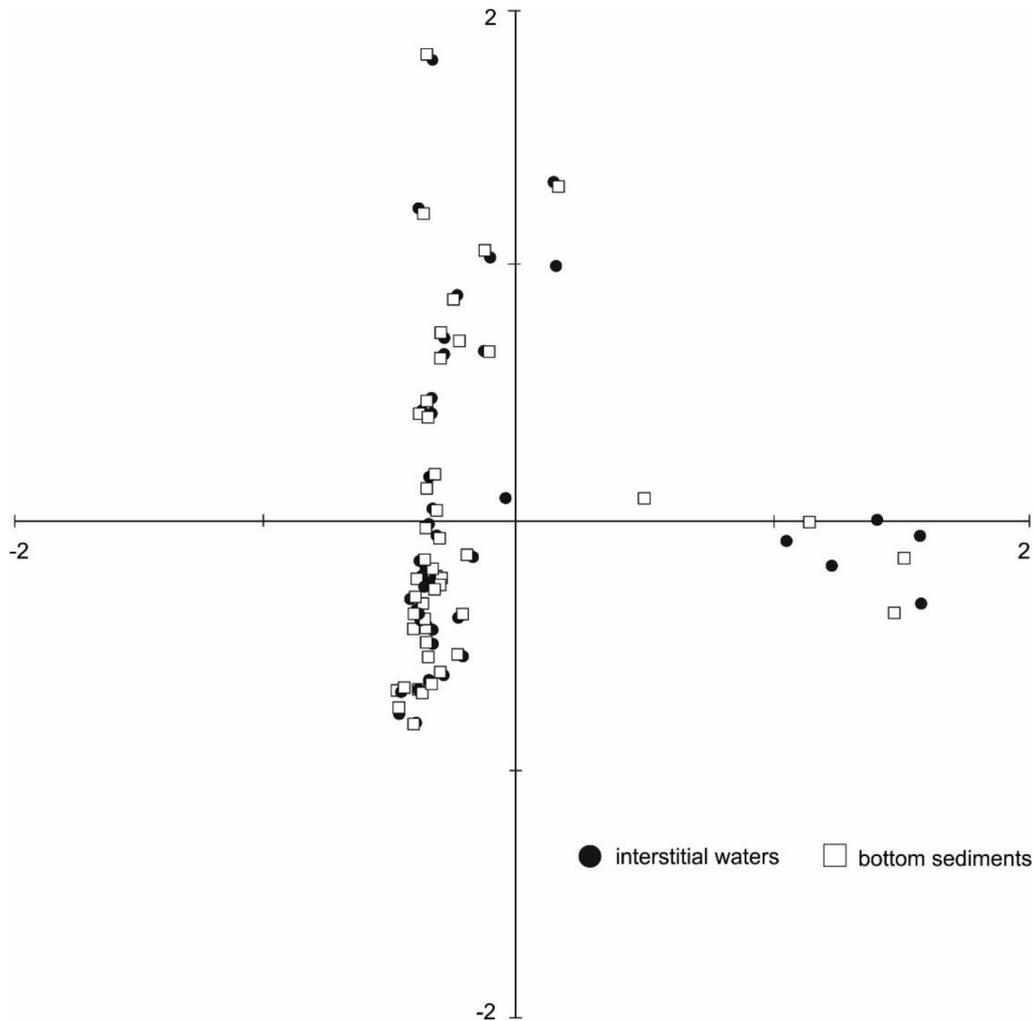


Fig. 4. Correspondence analysis (CA) ordination diagram with only plotted samples from interstitial waters and bottom sediments in the rivers.

conducted by Hajduk and Ogorzałek (1978), Sywula (1989), Dumnicka (2000), and Skalski (personal comm.) in this area. Stygobiontic species belonging to various taxonomic groups, not only annelids, are infrequently caught, so some species are exclusively known from type locality, such as *Trichodrilus spelaeus* (Moszyński, 1936), or from a few localities, like *Niphargus casimiriensis* (Skalski, 1980). Such a situation has been identified in other countries as well (Martin *et al.*, 2009; Giani *et al.*, 2011; Chertoprud *et al.*, 2016). Second, some rare species as *Hrabeiella periglandulata*, *Potamodrilus fluviatilis*, *Chaetogaster langi*, *Epirodrius pygmaeus* (Kahl and Pilipiuk, 2004), as well as endemic species, are found in singular localities in Polish subterranean waters. Third, many common benthic species are only accidentally discovered in underground habitats. In particular, this includes species feeding on algae (*e.g.*, many representatives of Naididae) or characteristic for lakes and lowland rivers with muddy bottoms (genus *Potamothrix*, *Psammoryctides barbatus*). Moreover, among typically terrestrial enchytraeids (from genera *Achaeta*, *Fridericia* or *Henlea*) some species have been found in a few localities in permanent or temporary subterranean water

bodies. The presence of soil dwelling species from that family was also identified in the aquatic underground habitat by other investigators (Giani *et al.*, 2011; Martinez-Ansemil *et al.*, 2016). Finally, a small number of records concerning lumbricids and leech species is caused by scarce occurrence of the first family (except for *Eiseniella tetraedra*) and a lack of Hirudinea in natural subterranean waters of Poland, however representatives of this last mentioned group were occasionally found in cave waters (Sket *et al.*, 2001).

Among the studied habitats, the highest richness of annelid fauna was identified in interstitial waters. A few factors seem to be responsible for this phenomenon. First, the highest number of records comes from this habitat. Furthermore, interstitial waters could be a refuge for various invertebrates during extreme hydrological events such as drought or flood (Wood *et al.*, 2010; Stubbington, 2012). Moreover, lower predator pressure and higher amounts of organic matter exist in interstitial waters compared to deeper subterranean waters (Williams *et al.*, 2010), facilitating survival of many benthic taxa (including oligochaetes) there. This is why their total number (Williams and Hynes, 1974; Olsen and Townsend, 2005)

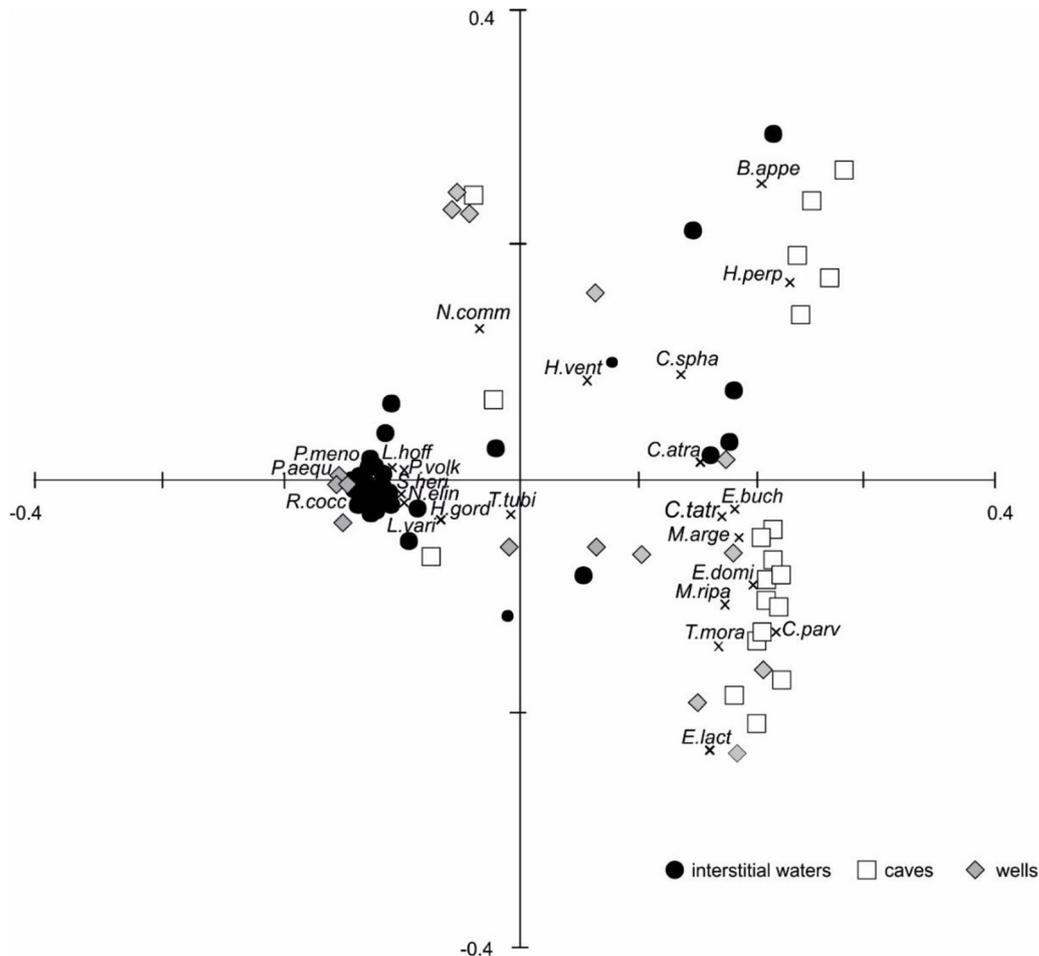


Fig. 5. Correspondence analysis (CA) ordination diagram with plotted samples and species of Oligochaeta from the studied underground water types. Abbreviations: B.appe- *Buchholzia appendiculata*, C.atra- *Cernosvitoviella atrata*, C.parv- *Cernosvitoviella parviseta*, C.tatr- *Cernosvitoviella tatrensis*, C.spha- *Cognettia sphagnetorum*, E.buch- *Enchytraeus buchholzi*, E.domi- *Enchytraeus dominicae*, E.lact- *Enchytraeus lacteus*, H.gord- *Haplotaxis gordioides*, H.perp- *Henlea perpusilla*, H.vent- *Henlea ventriculosa*, L.hoff- *Limnodrilus hoffmeisteri*, L.vari- *Lumbriculus variegatus*, M.arge- *Marionina argentea*, M.ripa- *Marionina riparia*, N.comm- *Nais communis*, N.elin- *Nais elinguis*, P. aequ- *Pristina aequisetata*, P.meno- *Pristina menoni*, P.volk- *Propappus volki*, R.cocc- *Rhyacodrilus coccineus*, S.heri- *Stylodrilus heringianus*, T. mora- *Trichodrilus moravicus*, T.tubi- *Tubifex tubifex*.

or frequency (Mary and Marmonier, 2000) could be higher than in benthos. In some Polish rivers, where annelid fauna was studied in both these habitats (Kasprzak, 1973a,b; Dumnicka, 2000), species richness was higher in interstitial waters, what confirms the importance of this habitat for benthic organisms.

The statistical analysis (CA) did not separate the samples of underground water types into distinct groups (Fig. 5), and the distribution of well-based fauna is especially scattered on the graph because abiotic parameters differ strongly in these habitats. In shallow open wells and in wells situated in river valleys, benthic taxa from various taxonomic groups were typically identified (Kasprzak, 1973a,b; Knight *et al.*, 2015; Dumnicka *et al.*, 2017), whereas in fairly deep wells supplied by infiltrating waters, annelids and crustaceans were mainly found (Dumnicka *et al.*, 2017). In general, benthic species also occurred in surface streams flowing through the caves, such as Skorocicka Cave (Dumnicka and Wojtan, 1993) or Wodna pod Pisaną Cave (Dumnicka and Galas, 1997) and in water bodies

situated near cave entrances (Chertoprud *et al.*, 2016), since it is easy for organisms to migrate and settle into these locations. However, in water bodies situated deep inside the caves, the number of oligochaete species was usually low, and stygobiontic species were found more often. Statistical analyses confirmed this for *Enchytraeus dominicae* only, because other stygobionts have only been caught sporadically, as mentioned above.

Aquatic and amphibious annelids species living on the bottom or in sediments are principally eyeless and detritivorous, so they are pre-adapted to subterranean life. Many species, especially those most frequently found in the underground habitat, could undergo their whole life cycle there, as indicated by findings of both juvenile and mature specimens or the chains of zooids of some Naididae species. These species belong to various families, including *Haplotaxis gordioides* (Haplotaxidae), *Stylodrilus heringianus*, *Lumbriculus variegatus* (Lumbriculidae), *Propappus volki* (Propap-

pidae), *Pristina aequisetata*, *P. menoni*, *Nais elinguis*, (Naididae), *Enchytraeus buchholzi*, and *Marionina argentea* (Enchytraeidae). These species are stygophiles; most of them prefer interstitial waters, but the latter two species were found mainly in caves. It is highly probable that some other enchytraeid taxa such as *Buchholzia appendiculata*, *Marionina riparia* and those representing the genus *Cernosvitoviella* are also stygophiles.

The results of available data analyses indicated that subterranean waters should be taken into account to recognize the whole diversity of annelids and other groups such as crustaceans and water mites, including subterranean waters that will prevent stygobionts from being overlooked. Moreover, investigations of the interstitial habitat should also be considered to obtain the composition of benthic invertebrates in running waters. Increasing human pressure on subterranean waters threatens the existence of stygobiontic and rare species; therefore, further studies on the distribution of these species are needed, and the subterranean diversity hot spots should be protected. In view of mounting threats faced by organisms living in this most widespread nonmarine environment on Earth, Mammola *et al.* (2019) advocated the necessity to promote in the general audience activities raising awareness about them.

References

- Chertoprud ES, Palatov DM, Borisov RR, Marinskiy VV, Bizin MS, Dbar RS. 2016. Distribution and a comparative analysis of the aquatic invertebrate fauna in caves of the western Caucasus. *Subterr Biol* 18: 49–70.
- Colwell RK. 2013. Estimates: Statistical estimation of species richness and shared species from samples. Version 9. User's Guide and application published at: <http://purl.oclc.org/estimates>
- Creuzé des Châtelliers P, Marmonier P, Dole–Oliver MJ, Castella E. 1992. Structure of interstitial assemblages in a regulated channel of the River Rhine (France). *Regulated Rivers Res Manag* 7: 23–30.
- Culver DC, Pipan T. 2009. The biology of caves and other subterranean habitats. Oxford: University Press.
- Culver DC, Pipan T. 2011. Redefining the extent of the aquatic subterranean biotope – shallow subterranean habitats. *Ecohydrology* 4: 721–730.
- Danielopol DL. 1984. Ecological investigations on the alluvial sediments of the Danube in the Vienna area – a phreatobiological project. *Verh Internat Verein Limnol* 22: 1755–1761.
- Demel K. 1918. Fauna jaskiń Ojcowskich. *Spraw Posiedzeń Tow Nauk Warsz Wydział Mat–Przyr* 11(4): 623–659.
- Dijkstra K-DB, Monaghan MT, Pauls SU. 2014. Freshwater biodiversity and insect diversification. *Ann Rev Entomol* 59: 143–163.
- Dumnicka E. 1996. Aquatic Oligochaeta and Aphanoneura from the souterrains of central Europe with description of a new *Enchytraeus* species. *Mém Biospéol* 23: 167–171.
- Dumnicka E. 2000. Studies on Oligochaeta taxocens in streams, interstitial and cave waters of southern Poland with remarks on Aphanoneura and Polychaeta distribution. *Acta zool cracov* 43: 339–392.
- Dumnicka E. 2003. Observations on the distribution of aquatic fauna in Tatra Mountain caves. *Subterr Biol* 1: 49–55.
- Dumnicka E. 2009. Diversity of oligochaete fauna inhabiting epigeal and hypogean sectors of the stream flowing through a non-karstic cave in Beskidy Mts. (Poland). *Subterr Biol* 7: 41–46.
- Dumnicka E, Galas J. 1997. The relationship between Oligochaeta, particulate organic matter and environmental conditions in epigeal and hypogean parts of a mountain stream in Poland. *Mém Biospéol* 24: 9–14.
- Dumnicka E, Galas J. 2017. An overview of stygobiontic invertebrates of Poland based on published data. *Subterr Biol* 23: 1–18.
- Dumnicka E, Wojtan K. 1993. Invertebrates (with special regard to Oligochaeta) of the semi-underground water bodies in the gypsum caves. *Mém Biospéol* 20: 63–67.
- Dumnicka E, Galas J, Karlikowska J, Sznober N. 2015. Temporary co-existence of aquatic and terrestrial invertebrates in shallow periodically flooded and frozen cave. *Biologia* 70: 1201–1209.
- Dumnicka E, Chabrowska A, Szokalski M. 2016. Fauna bezkręgowców w wypływie z Jaskini w Lelowie (Wyżyna Małopolska). *Ziemia Częstochowska* 42: 145–156.
- Dumnicka E, Galas J, Krodkiwski M. 2017. Patterns of benthic fauna distribution in wells: the role of anthropogenic impact and geology. *Vadose Zone J* 16: 1–9.
- Dumnicka E, Konopacka A, Zurek R. 2018. Changes in the benthic fauna composition in the Upper Vistula over the last 50 years – the consequences of the water pollution reduction and alien species invasion. *Oceanol Hydrobiol Stud* 47: 303–312.
- Giani N, Sambugar B, Martinez-Ansemil E, Martin P, Schmelz RM. 2011. The groundwater oligochaetes (Annelida, Clitellata) of Slovenia. *Subterr Biol* 9: 85–102.
- Gibert J, Danielopol DL, Stanford JA. 1994a. Groundwater ecology. London: Academic Press.
- Gibert J, Stanford JA, Dole-Olivier M-J, Ward JV. 1994b. Basic attributes of groundwater ecosystems and prospects for research. In: Gibert J, Danielopol DL, Stanford JA. Eds. Groundwater ecology. London: Academic Press, 7–40.
- Hajduk Z, Ogorzałek A. 1978. *Niphargellus arndti* (Schellenberg, 1933) z jaskini Kontaktowej koło Kletna. *Acta Univ Wratisl 3. Studia Geograficzne* 24: 155–157.
- Jaworowski A. 1893. Fauna studzienna miast Krakowa i Lwowa. *Spraw Kom Fizyograf AU w Krakowie* 28: 29–48.
- Juberthie Ch, Decu V. (eds) 1994. Encyclopedia Biospeologica, vol. 1. Bucarest: Société de Biospéologie Moulis.
- Kahl K, Pilipiuk I. 2004. Skąposzczety (Oligochaeta) In: Bogdanowicz W, Chudzicka E, Pilipiuk I, Skibińska E. eds. Fauna of Poland Characteristics and checklist of species. Warszawa: Muzeum i Instytut Zoologii PAN, pp. 9–17.
- Kasprzak K. 1973a. Notatki o faunie skąposzczetów (Oligochaeta) Polski I. *Fragm Faun* 18: 405–434.
- Kasprzak K. 1973b. Notatki o faunie skąposzczetów (Oligochaeta) Polski II. *Fragm Faun* 19: 1–19.
- Kasprzak K. 1973c. Skąposzczety (Oligochaeta) wód interstycjalnych. *Przeł Zool* 17: 41–44.
- Kasprzak K. 1975. Skąposzczety (Oligochaeta) osadów aluwialnych rzek. *Prace Komisji Naukowych PTG III/16*: 173–185.
- Kasprzak K. 1976. Badania nad skąposzczetami (Oligochaeta) dolnego biegu rzeki Wełny. *Fragm Faun* 20: 425–466.
- Kasprzak K. 1979a. Skąposzczety (Oligochaeta) Pienin. I. Wazonkowce (Enchytraeidae). *Fragm Faun* 24: 7–56.
- Kasprzak K. 1979b. Skąposzczety (Oligochaeta) Pienin. Naididae, Tubificidae, Haplataxidae, Lumbriculidae, Branchiobdellidae. *Fragm Faun* 24: 57–80.
- Kasprzak K, Zajonc I. 1980. Skąposzczety (Oligochaeta) Tatr. *Przeł Zool* 24: 189–199.

- Knight LRFD, Brancelj A, Hänfling B, Cheney C. 2015. The groundwater invertebrate fauna of the Channel Islands. *Subterr Biol* 15: 69–94.
- Košel V. 2009. Subterranean fauna Zapadnych Karpat. Czech Academy of Sciences: České Budějovice.
- Krzanowski W, Fiedor E, Kuflikowski T. 1965. Fauna denna kamienisto-prądowych siedlisk dolnych odcinków Białego Dunajca, Rogoźnika i Lepietnicy. *Zesz Nauk UJ 103, Prace Zoologiczne* 9: 43–60.
- Łuczak J, Rybak M, Ranke-Rybicka B. 1980. Występowanie organizmów wodnych w wodzie wodociągowej. *Rocznik PZH* 31: 319–325.
- Mammola S, Cardoso P, Culver DC, *et al.* 2019. Scientists' Warning on the Conservation of Subterranean Ecosystems. *BioScience* 69: 641–650.
- Martin P, De Broyer C, Fiers F, Michel G, Sablon R, Wouters K. 2009. Biodiversity of Belgian groundwater fauna in relation to environmental conditions. *Freshwat Biol* 54: 814–829.
- Martínez-Ansemil E, Giacomazzi F, Sambugar B. 2016. Groundwater oligochaetes (Annelida: Clitellata) of the Dinaric region (South-East Europe). *Biologia* 71: 24–30.
- Mary N, Marmonier P. 2000. First survey of interstitial fauna in New Caledonian rivers: influence of geological and geomorphological characteristics. *Hydrobiologia* 418: 199–208.
- Meleg I, Campean M, Pavelescu C. 2009. Hyporheic fauna from the interstitial of the Somes river basin (Transylvania, Northwestern Romania). *Trav Inst Spéol "Emile Racovitza"* 48: 45–58.
- Moszyński A. 1934. Skąposzczety (Oligochaeta) miasta Poznania. *Kosmos ser. A* 57: 235–255.
- Moszyński A. 1936. Die Höhlenfauna des Glatzer Schneeberges. 9. Ein neuer Vertreter der Gattung *Trichodrilus* Clap. (*Trichodrilus spelaeus* nov. spec.) aus dem Stollen in Neu-Klessengrund. *Beitr Biol Glatz Schneeberges* 2: 214–216.
- Olsen DA, Townsend CR. 2005. Flood effects on invertebrates, sediments and particulate organic matter in the hyporheic zone of a gravel-bed stream. *Freshwat Biol* 50: 839–853.
- Ortuño VM, Gilgado JD, Jiménez-Valverde A, Sendra A, Pérez-Suárez G, Herrero-Borgoñón JJ. 2013. The "alluvial mesovoid shallow substratum" a new subterranean habitat. *Plos One* 8: e76311.
- Pacioglu O. 2010. Ecology of the hyporheic zone: a review. *Cave and Karst Science* 36: 69–76.
- Pawłowski LK. 1951. Pijawki (Hirudinaea) stacji pomp rzecznych oraz stacji filtrów w Warszawie. *Fragm Faun Mus Zool Pol* 6: 169–192.
- Pax F, Maschke K. 1935. Die Höhlenfauna des Glatzer Schneeberges. 1. Die rezente Metazoenfauna. *Beitr Biol Glatz Schneeberges* 1: 4–72.
- Skalski A. 1967. Characteristics of the recent fauna from the Szczelina Chochołowska cave in the Tatra Mts. *Prace Muzeum Ziemi* 11: 288–290.
- Skalski AW. 1980. Studniczek Iwowski, *Niphargus leopoliensis* Jaworowski, 1893 (Amphipoda) w Polsce. *Przeegl Zool* 24: 97–101.
- Sket B. 1999. High biodiversity in hypogean waters and its endangerment – the situation in Slovenia, the Dinarctic Karst, and Europe. *Crustaceana* 72: 767–779.
- Sket B, Dovč P, Jalžić B, Kerovec M, Kučinić M, Trontelj P. 2001. A cave leech (Hirudinea, Erpobdellidae) from Croatia with unique morphological features. *Zool Scr* 30: 223–229.
- Stammer H-J. 1936. Die Höhlenfauna des Glatzer Schneeberges. 8. Die Wasserfauna der Schneeberghöhlen. *Beitr Biol Glatz Schneeberges* 2: 199–214.
- Starmach K, Wróbel S, Pasternak K. 1976. *Hydrobiologia. Limnologia*. Państwowe Wydawnictwo Naukowe, Warszawa.
- Stubbington R. 2012. The hyporheic zone as an invertebrate refuge: a review of variability in space, time, taxa and behaviour. *Mar Freshw Res* 63: 293–311.
- Sywula T. 1989. *Bathynella natans* Vejdovsky, 1882 i *Proasellus slavus* (Remy, 1948) nowe dla Polski, podziemne skorupiaki. *Przeegl Zool* 33: 77–82.
- Ter Braak CJF, Šmilauer P. 2002. CANOCO reference manual and CanoDraw for windows user's guide: Software for canonical community ordination (version 4.5), Microcomputer Power Ithaca, NY, USA
- Williams DD, Febria CM, Wong JCY. 2010. Ecotonal and other properties of the hyporheic zone. *Fundam Appl Limnol, Arch Hydrobiol* 176: 349–364.
- Williams DD, Hynes HBN. 1974. The occurrence of benthos deep in the substratum of a stream. *Freshw Biol* 4: 233–256.
- Wood PJ, Boulton AJ, Little S, Stubbington R. 2010. Is the hyporheic zone a refugium for aquatic macroinvertebrates during severe low flow conditions? *Fundam Appl Limnol, Arch Hydrobiol* 176: 377–390.

Cite this article as: Dumnicka E, Galas J, Krodkiewska M, Pocięcha A. 2020. The diversity of annelids in subterranean waters: a case study from Poland. *Knowl. Manag. Aquat. Ecosyst.*, 421, 16.